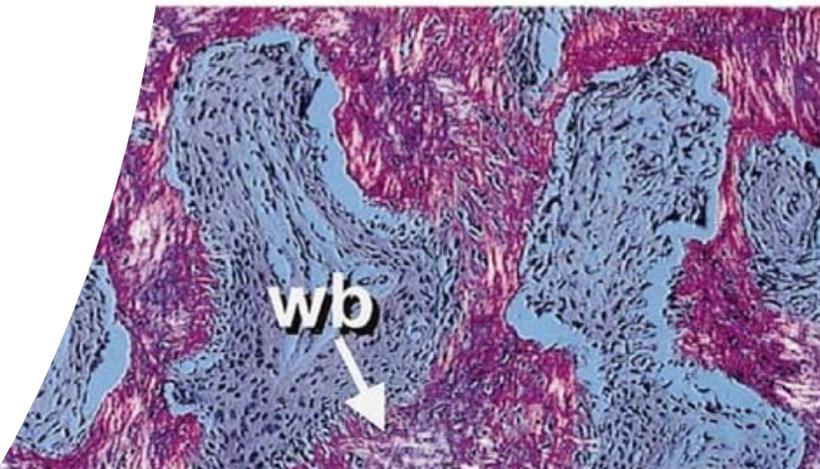
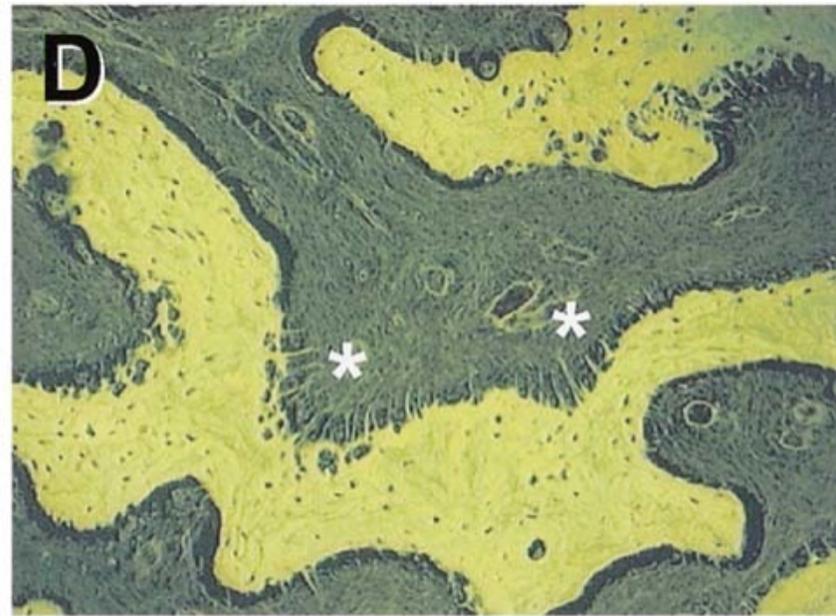
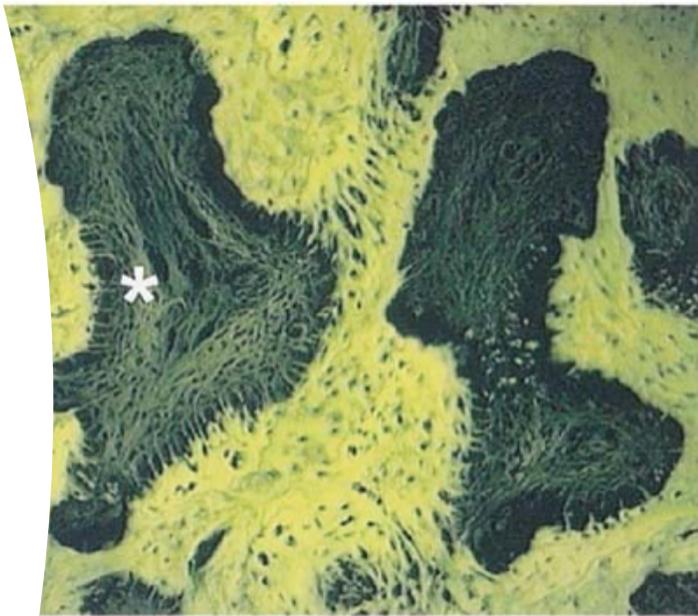
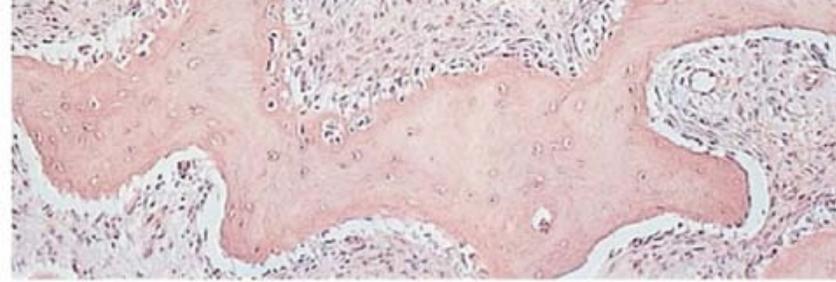
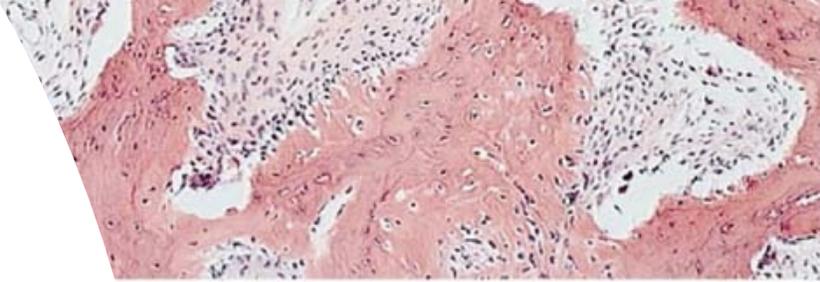
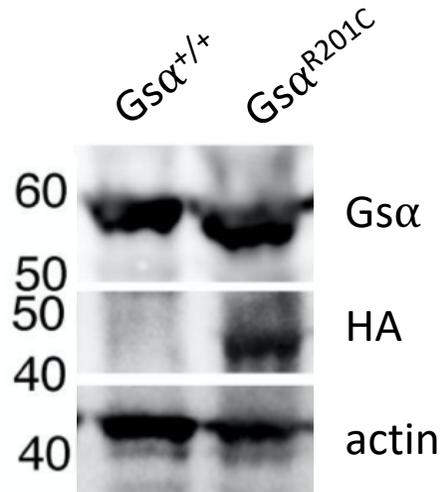


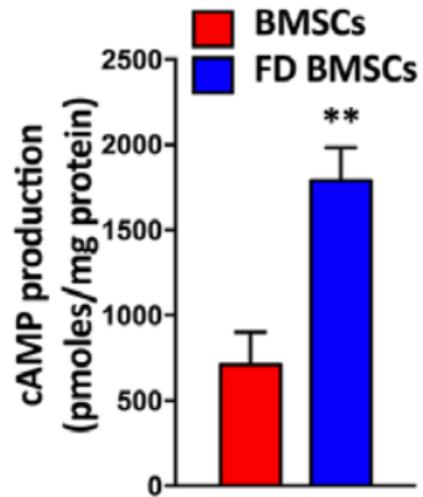
GNAS and HDAC8:
potential therapeutic targets
to restore differentiation
of BMSCs
in Fibrous Dysplasia



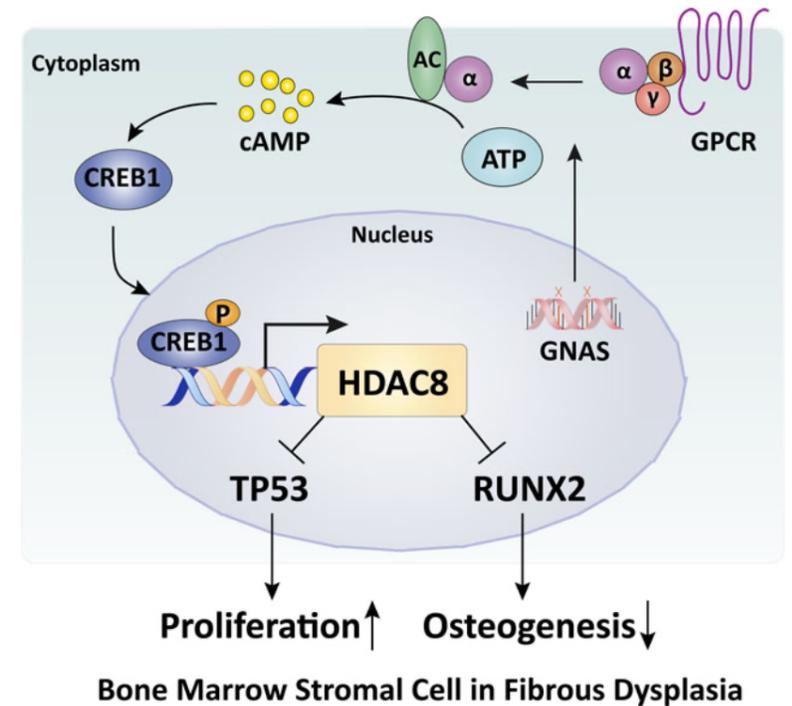
Role of GNAS and HDAC8 in Fibrous Dysplasia



Adapted by: Raimondo et al., 2020



Xiao, Fu, Zhu et al., 2018



Xiao, Fu, Zhu et al., 2018

FIRST AIM

Use of CRISPR/Cas13 on HDAC8 to interfere with HDAC8 activity in FD BMSCs



Reduce HDAC8 activity to restore physiological osteogenic differentiation

SECOND AIM

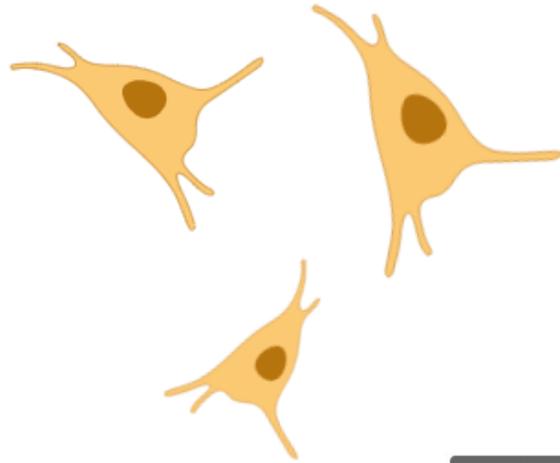
Use of CRISPR/Cas9 on GNAS mutated gene in human FD BMSCs



Restore WT Gs α phenotype

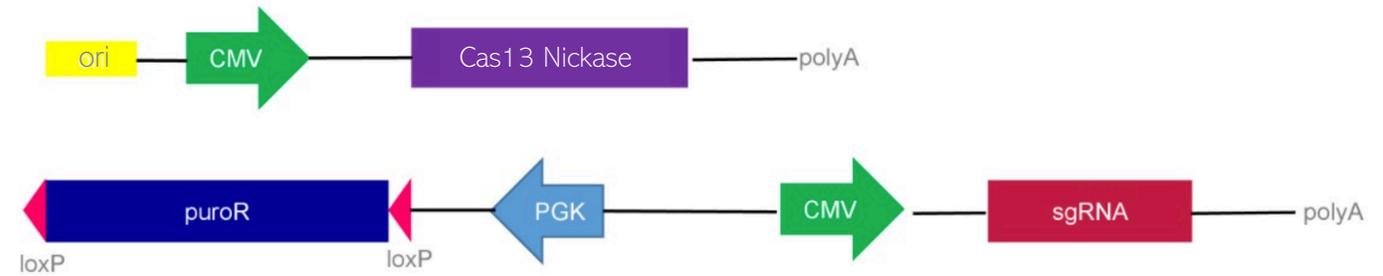
CRISPR/Cas13 on HDAC8 – Material

Human FD CD146/MCAM cells



Created in BioRender.com bit

Donor vector
pDEST-IRES-puro-polyA



HDAC8 sgRNA

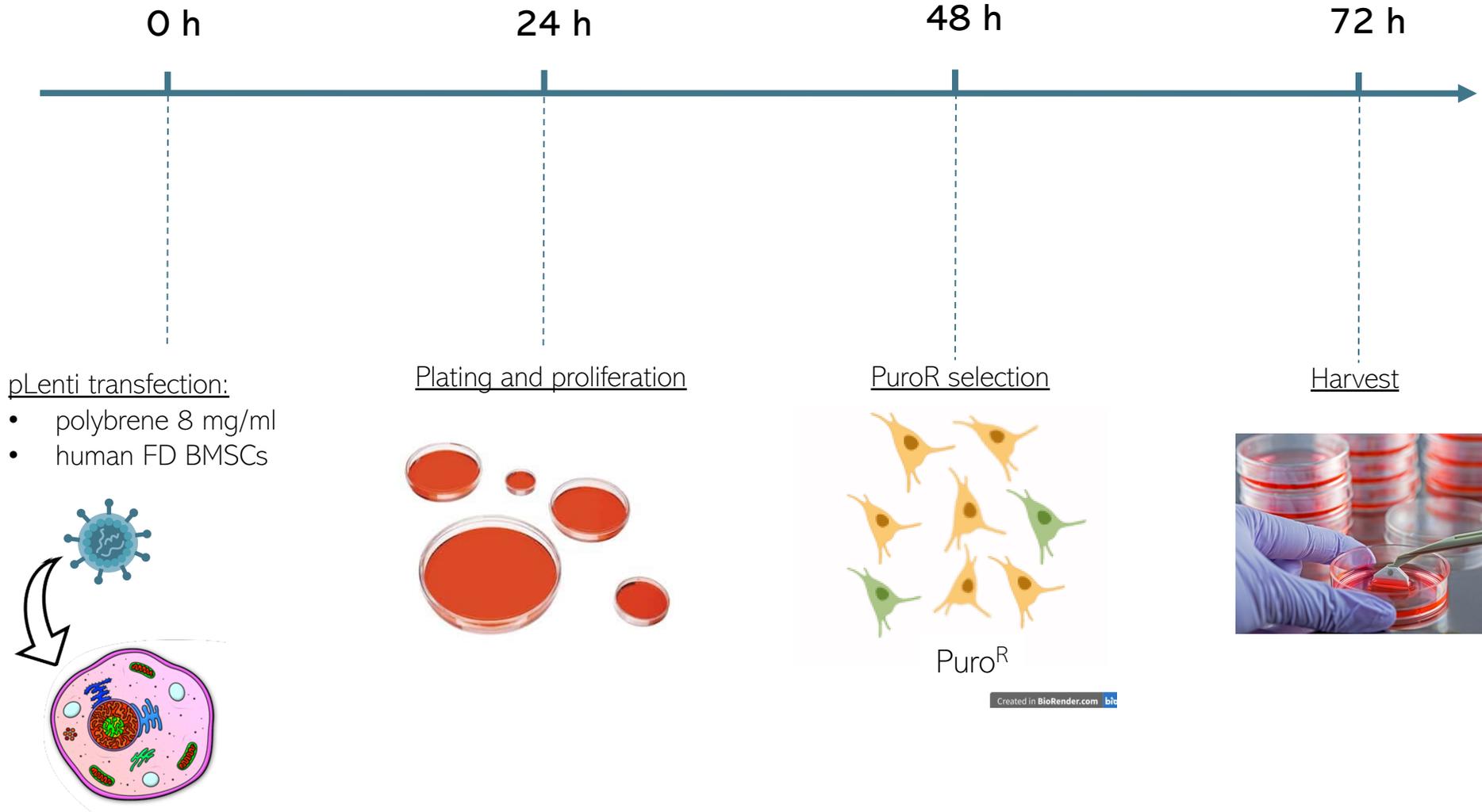
- 1: 5-GACGTGTCTGATGTTGGCCTNGG-3
- 2: 5-GCGGAAGATGGAGGAGCCGGNGG-3
- 3: 5-GTAGCAATTA ACTGGTCTGGNGG-3

<http://www.e-crisp.org>

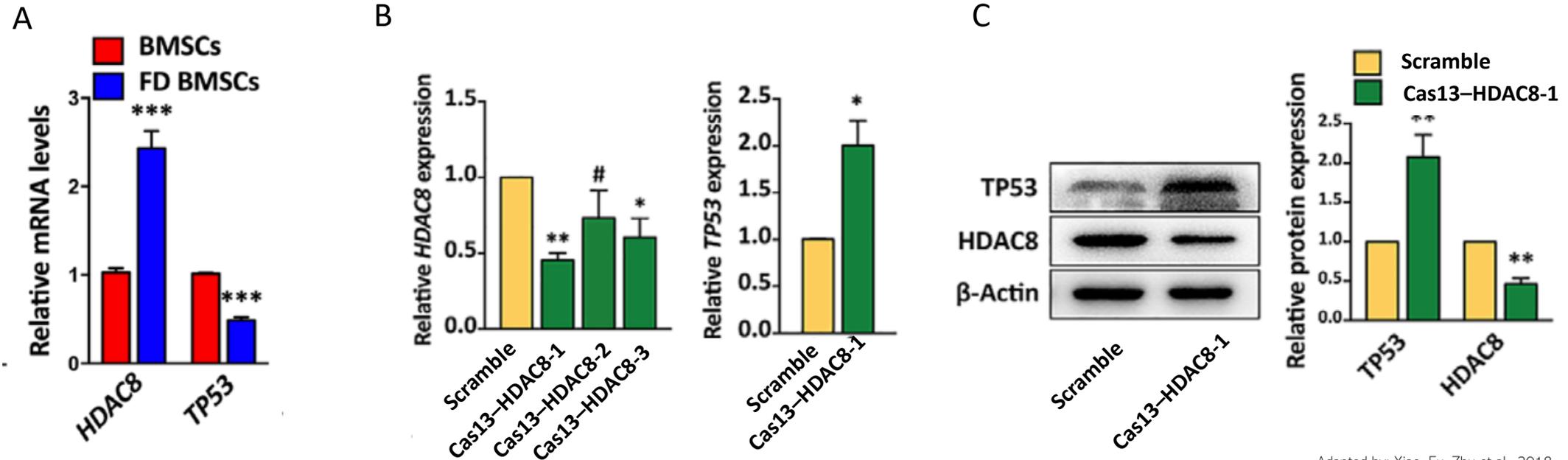
CRISPR/Cas13 on HDAC8 – Methods

Incubation at 37°C:

- 25 cm² plastic flasks
- DMEM medium
- 100 U/ml penicillin
- 100 µg/ml streptomycin
- 10% fetal bovine serum



CRISPR/Cas13 on HDAC8 – *In vitro* results I



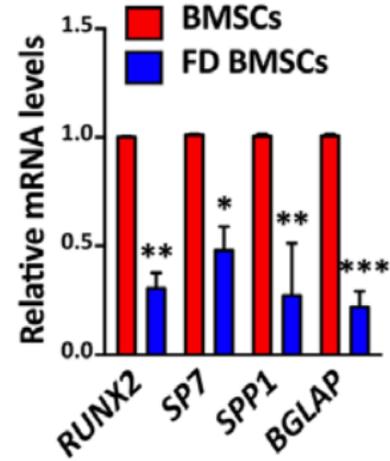
Adapted by: Xiao, Fu, Zhu et al., 2018

Graphic A. RT-PCR showing HDAC8 and TP53 mRNA level expression after FD BMSCs were cultured in osteogenesis induction medium for 7 days.

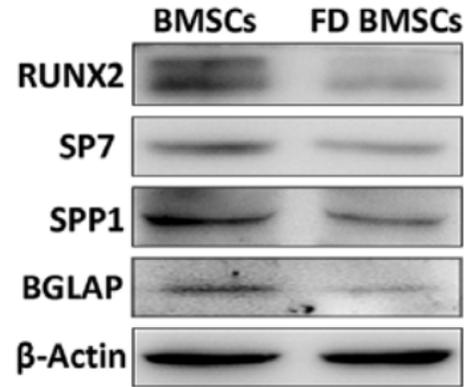
Graphic B-C. RT-PCR and western blot of HDAC8 and TP53 expression in FD BMSCs with CRISPR/Cas13 compared with the control.

CRISPR/Cas13 on HDAC8 – *In vitro* results II

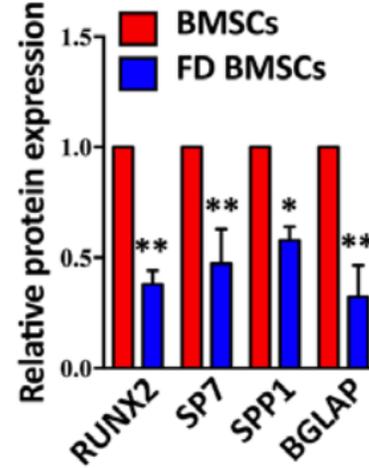
A



B

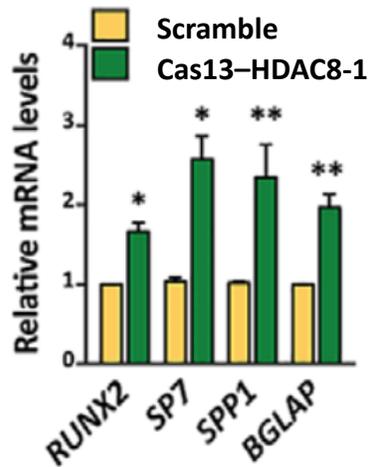


C

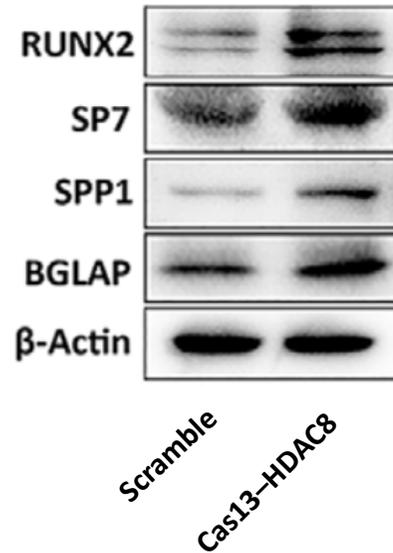


Graphics A-C. 1: RT-PCR showing osteogenesis marker mRNA expression. 2-3: western blot analysis of osteogenesis markers. Data collected from FD BMSCs in osteogenesis induction medium for 7 days.

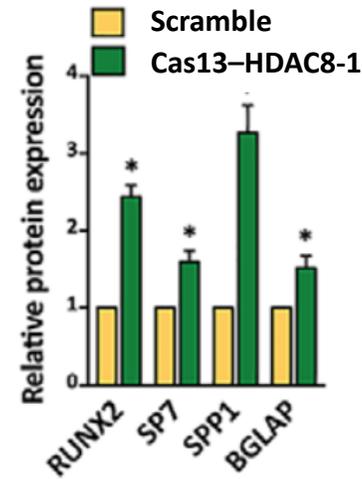
D



E

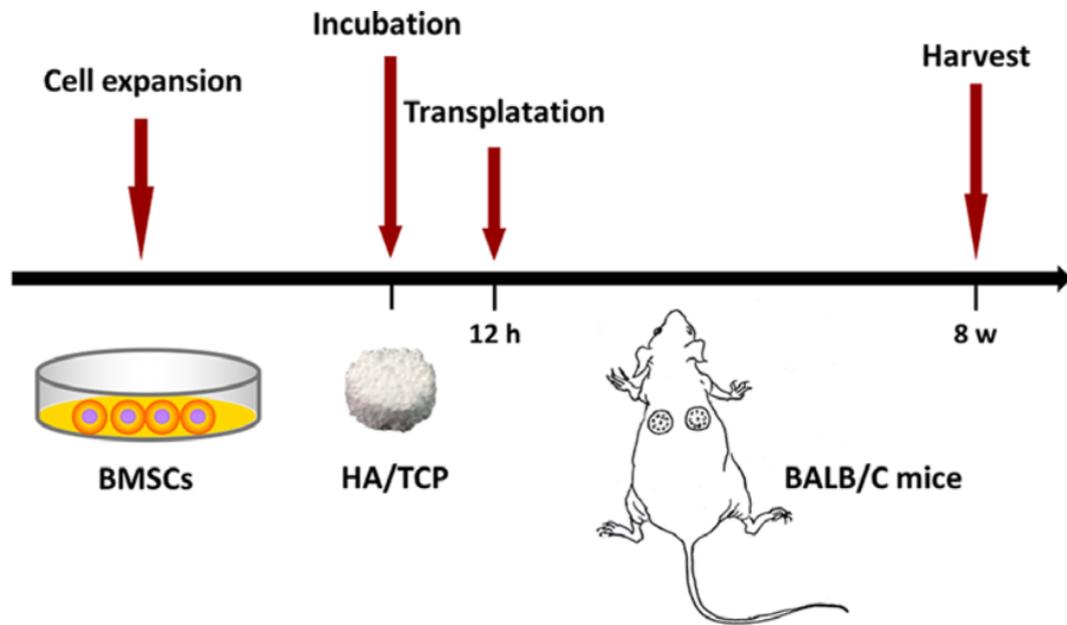


F

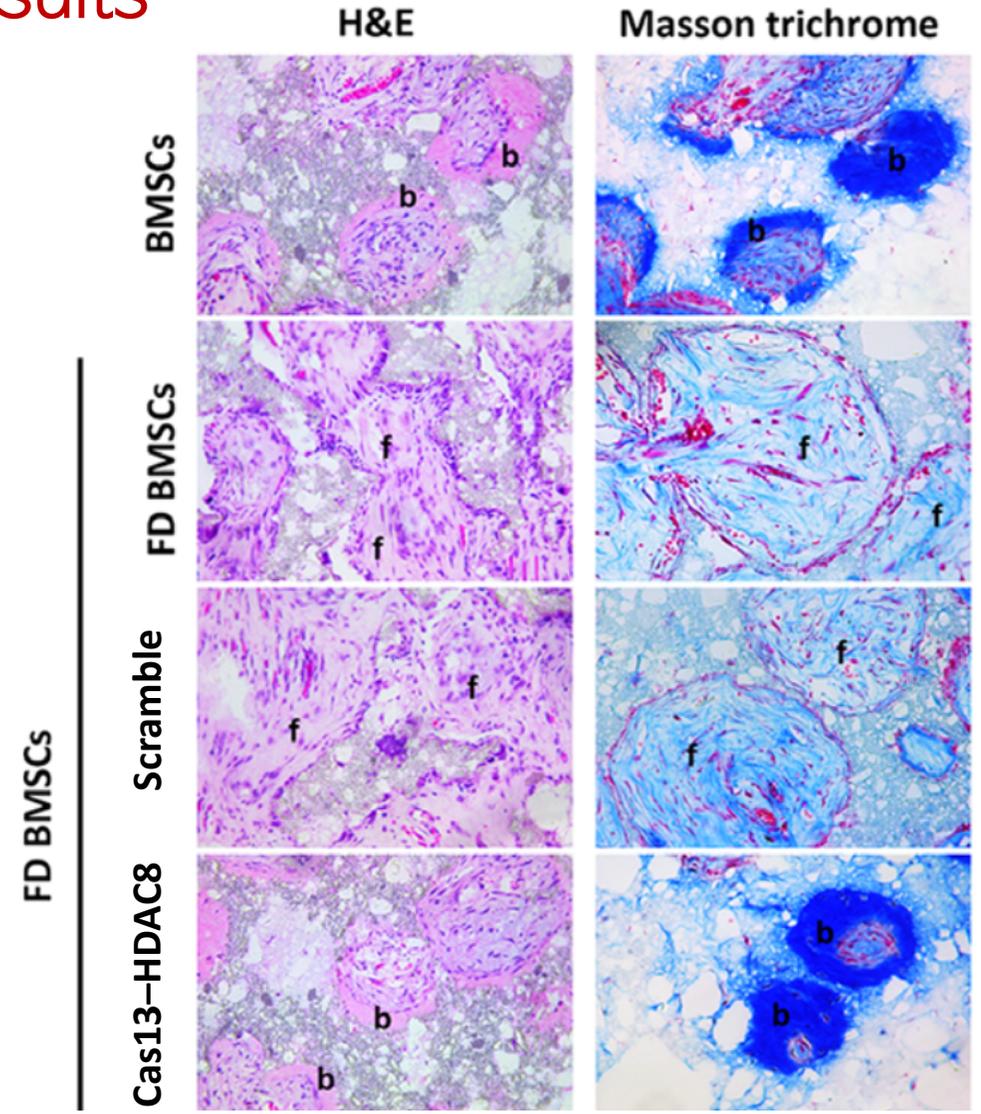


Graphics D-F. RT-PCR and western blot of osteogenesis marker expression in FD BMSCs with CRISPR/Cas13 compared with the control.

CRISPR/Cas13 on HDAC8 – *In vivo* results



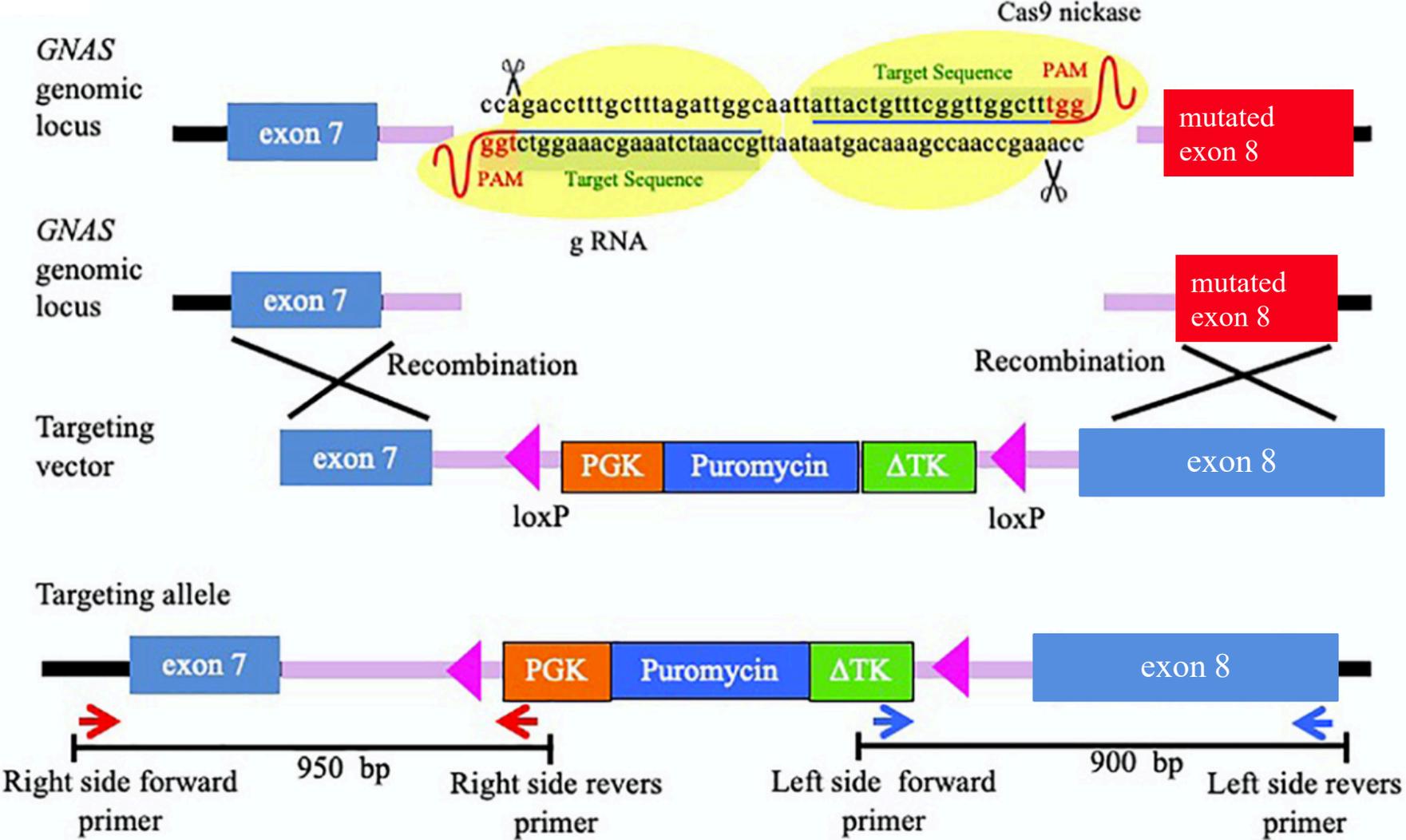
Adapted by: Xiao, Fu, Zhu et al., 2018



Adapted by: Xiao, Fu, Zhu et al., 2018

b = bone tissue
ft = fibrous tissue

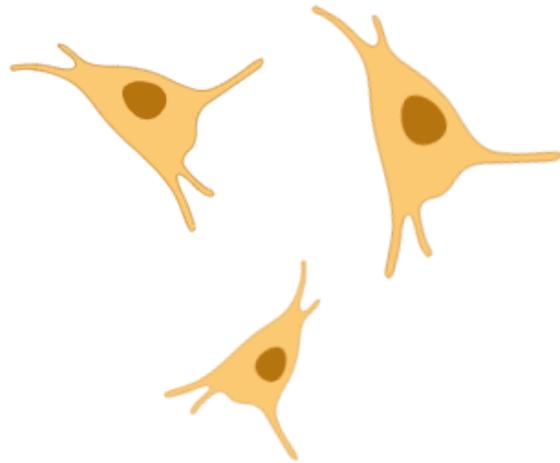
CRISPR/Cas9 strategy – Material and Methods



adapted by: Watanabe et al., 2020

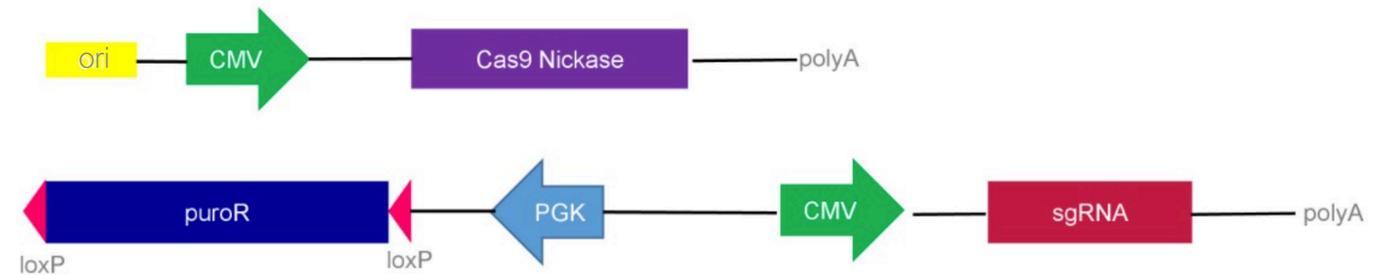
CRISPR/Cas9 on GNAS – Material and methods

Human FD CD146/MCAM cells



Created in BioRender.com

Donor vector
pDEST-IRES-puro-polyA

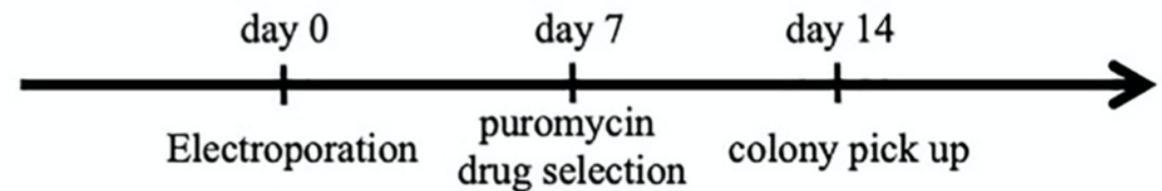


GNAS gsRNA

1: 5-GAGGCGATTGAAGTACGTGCNGG-3

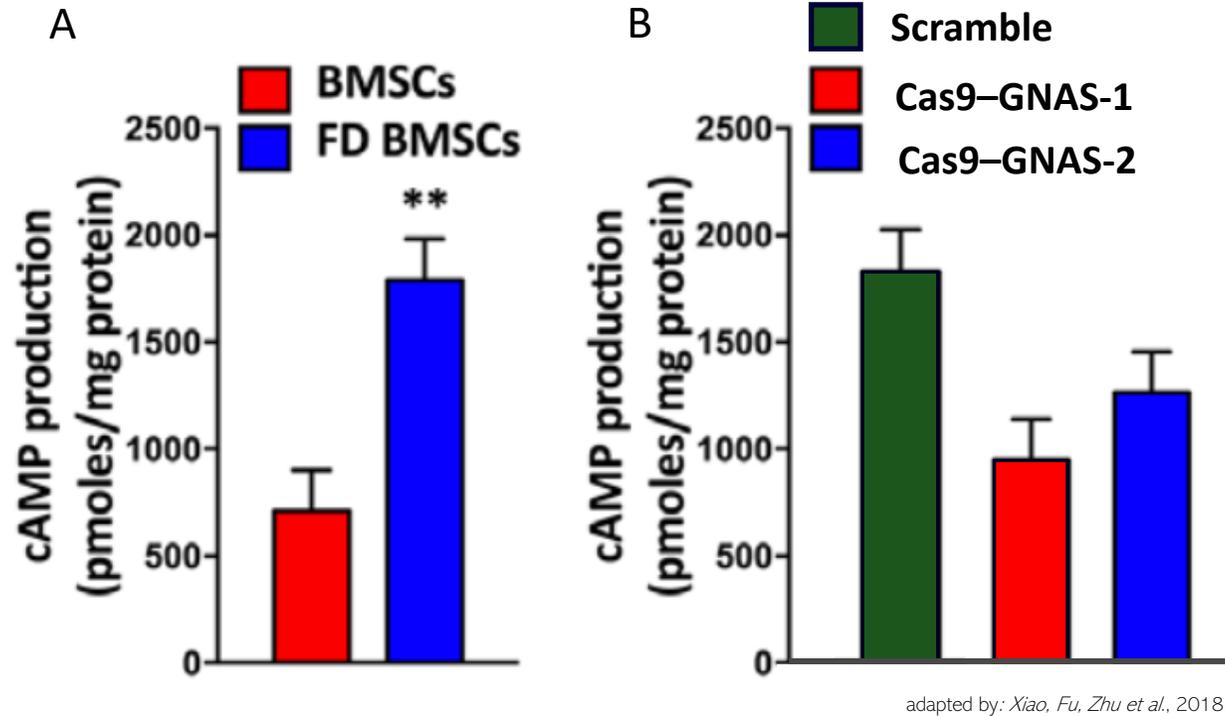
2: 5-GCTGCTTCTAGGTAATGCGGNGG-3

<http://www.e-crisp.org>

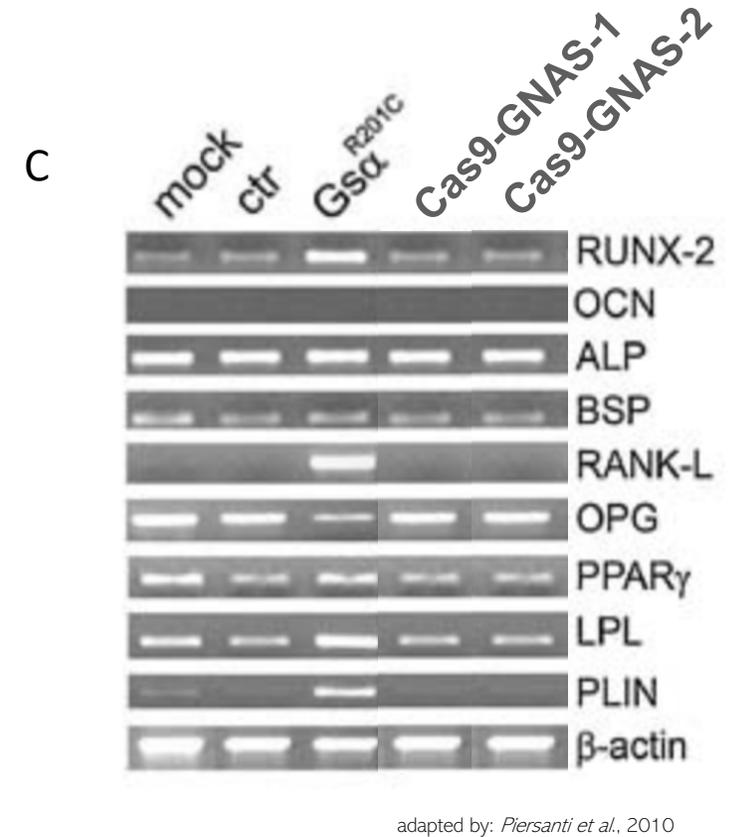


adapted by: *Watanabe et al.*, 2020

CRISPR/Cas9 strategy - *In vitro* results



Graphic A-B. cAMP levels in FD BMSCs and control cells analyzed by ELISA .



Graphic C. RT-PCR showing osteogenic marker quantification in Gsα^{WT}-transduced BMSCs.

CRISPR/Cas9 strategy - *In vivo* transplant expectations



Scramble BMSCs



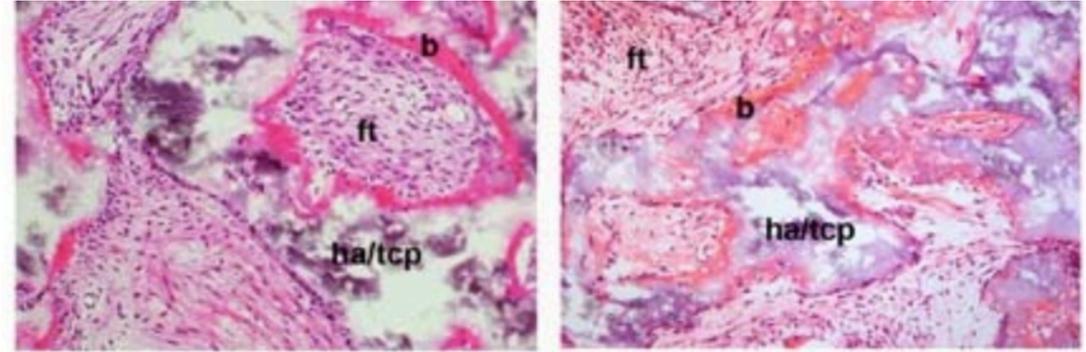
Cas9-GNAS-1 BMSCs



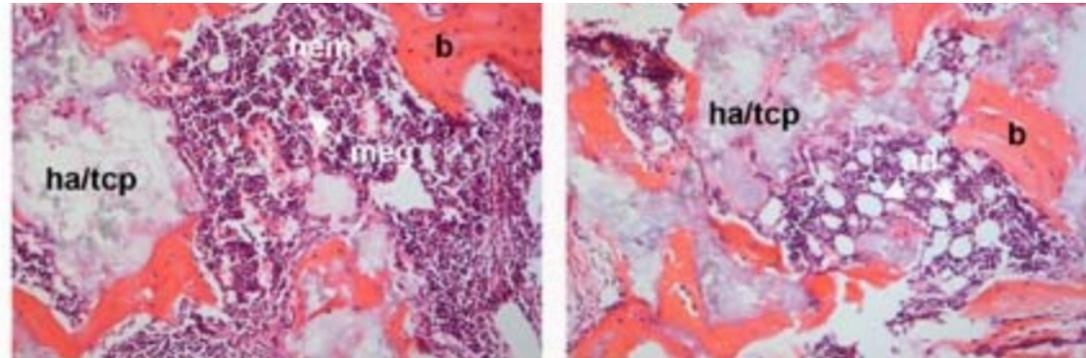
HA intraosseal

Created in BioRender.com

Scramble BMSCs



Cas9-GNAS-1 BMSCs



adapted by: Piersanti et al., 2010

b = formation of abundant bone
ft = fibrous tissue
ha/tcp = hydroxyapatite/tricalcium phosphate
hem, meg = megakaryocytes
ad = adypocytes

Conclusions

CRISPR/CAS13 STRATEGY

- *In vitro* collected data show restoration of normal mRNA and protein expression levels in FD BMSCs transfected with CRISPR/Cas13 donor vector;
- *In vivo* treatment in mice shows normal bone tissue development, resulting in a WT-like phenotype;
- These results suggest that HDAC8 could be a potential hopeful therapeutic target to fight Fibrous Dysplasia in BMSCs.

CRISPR/CAS9 STRATEGY

- Innovative CRISPR technology allows to correct specifically the GNAS mutation to effectively heal Fibrous Dysplasia;
- *In vitro* experiment demonstrates restoration of cAMP normal level, resulting in a WT-like expression of osteogenic markers;
- *In vivo* experiments showed ossicles formation in treated mice.

Costs

MATERIALS

| | |
|--|-------------------|
| •12X lentiCRISPR v2 Addgene | € 952.30 |
| •16X 3-weeks-old male BALB/C mice Jax | € 271.00 |
| •1X QIAGEN OneStep RT-PCR Kit (100) Qiagen | € 519.00 |
| •2X QIAquick PCR Purification Kit (50) Qiagen | € 208.00 |
| •3X RUNX2 Western blot kit (AWBK41328) Avasysbio | € 1,394.00 |
| •2L DMEM – Dulbecco’s modified Eagle Medium Thermofischer | € 76.00 |
| •10 mg Puromycin Sigmaaldrich | € 63.50 |
| •1X Trichrome Stain Kit (ab150686) Abcam | € 312.26 |
| •1X H&E Staining Kit (ab245880) Abcam | € 131.20 |
| TOTAL | € 3,927.26 |

STAFF

| | |
|--------------------------|---------------------|
| •1X Class V Researcher | € 60,084.02 |
| •2X Class III Researcher | € 109,736.68 |
| •1X Post PhD student | € 19,012.76 |
| •1X PhD student | € 16,067.88 |
| •1X Animal technician | € 28,500.00 |
| TOTAL per Year | € 233,401.34 |



References

- *Adli*, The CRISPR tool kit for genome editing and beyond. Nature Communications 2018; 9:1911
- *Piersanti et al.* Transfer, analysis, and reversion of the fibrous dysplasia cellular phenotype in human skeletal progenitors. J Bone Miner Res. 2010; 25(5):1103-16.
- *Raimondo et al.* Changes in gene expression in human skeletal stem cells transduced with constitutively active Gs α correlates with hallmark histopathological changes seen in fibrous dysplastic bone. PLoS ONE 2020; 15(1): e0227279.
- *Saggio*. Perils and Promises of Therapeutic Approaches for the Stem Cell Disease Fibrous Dysplasia. Stem Cells Translational Medicine 2019; 8:110–111
- *Watanabe et al.* A novel GNAS-mutated human induced pluripotent stem cell model for understanding GNAS-mutated tumors. Tumor biology 2020; Sep: 1 – 13.
- *Xiao, Fu, Zhu et al.* HDAC8, A Potential Therapeutic Target, Regulates Proliferation and Differentiation of Bone Marrow Stromal Cells in Fibrous Dysplasia. Stem Cells Translational Medicine 2019; 8:148–161.
- *Yu Fu et al.* Histone deacetylase 8 suppresses osteogenic differentiation of bone marrow stromal cells by inhibiting histone H3K9 acetylation and RUNX2 activity. The International Journal of Biochemistry & Cell Biology (2014); 54:68–77.