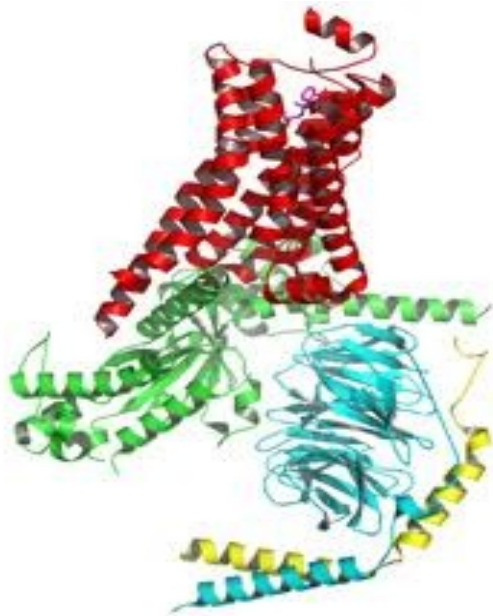




SAPIENZA  
UNIVERSITÀ DI ROMA

Corso Terapia Genica  
AA 2012/13  
Prof.ssa Isabella Saggio

# POH: new model for Gene Therapy using Lentivectors



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# Progressive Osseous Heteroplasia (POH)

## Genetic disorder of Mesenchymal differentiation:

- dermal ossification during infancy and progressive heterotopic ossification of cutaneous, subcutaneous, and deep connective tissues during childhood.

## Autosomal dominant disorder:

- heterozygous mutations in *GNAS1* (encoding the  $\alpha$ -subunit of the stimulatory G protein of adenylyl cyclase)
- altered regulation of cyclic AMP-mediated signal transduction in MSC

# Progressive Osseous Heteroplasia (POH)

## Clinical features of heterotopic ossification in POH

<i>Feature</i>	<i>POH</i>
Sex distribution	Female = male
Genetic transmission	Autosomal dominant
Congenital malformation of great toes	-
Congenital papular rash	+
Cutaneous ossification	+
Subcutaneous ossification	+
Muscle ossification	+
Superficial to deep progression of ossification	+
Severe limitation of mobility	+
Severe flare-ups of disease	-
Ectopic ossification after intramuscular injections	-
Ectopic ossification after trauma	±
Regional patterns of progression	-
Definitive treatment available	-

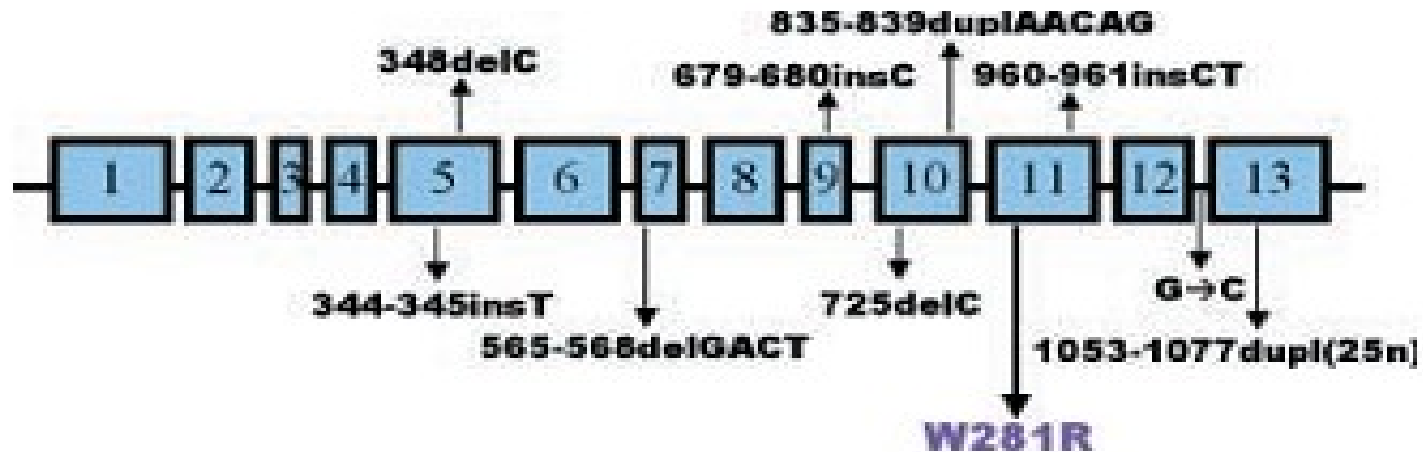
## Pathological and laboratory features

<i>Feature</i>	<i>POH</i>
<b>Predominant mechanism of ossification</b>	<b>Intramembranous</b>
Inflammatory perivascular and muscle infiltrate	-
Hematopoietic marrow in ectopic bone	±
PTH resistance	-
Hypocalcemia and hyperphosphatemia	-
<b>Pathogenesis</b>	<b>Unknown</b>

Kaplan and Shore, (2000)

# Progressive Osseous Heteroplasia (POH)

## MUTATIONS

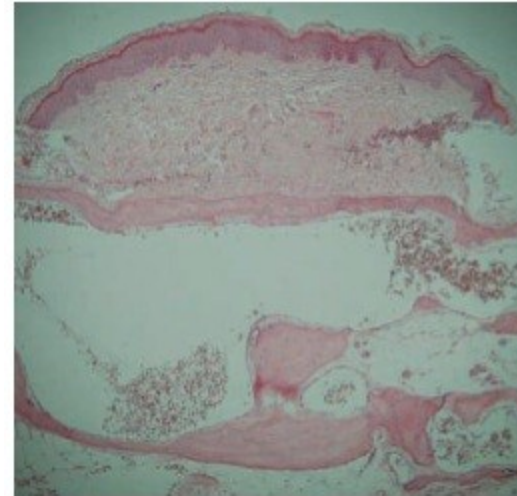


**Figure 4** Illustration of the positions of the previously described heterozygous inactivating mutations identified in the *GNAS1* gene in patients with progressive osseous heteroplasia<sup>2</sup> and the new missense mutation described here (W281R).

# Progressive Osseous Heteroplasia (POH)



Leg of patient aged 9 years old



Lesion on the leg shows mature bone formation in the dermis (haematoxylin & eosin)

# Preliminary literatures dates

## NO CURE FOR POH

- Surgery: highly discouraged;
- In vitro studies:  
BMSC (bone marrow stromal cells)  
STSC (soft tissue stromal cells-adipose);
- In vivo models does not exist.

# GNAS locus

## Complex locus:

- contains **independent imprinting** domains that **uses multiple promoters** to generate several gene products
- 20q13.2-13.3 in the human genome

## Encodes to 4 different isoforms of Gs- $\alpha$ Proteins:

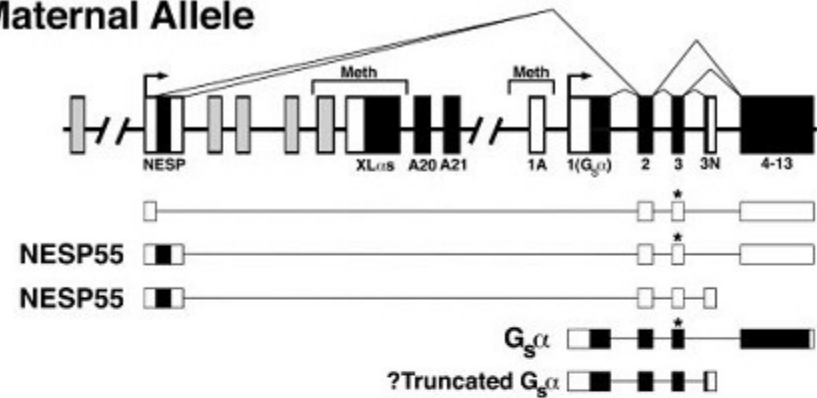
- two longs (Gsa-1 and 2)
- two shorts (Gsa-3 and 4)

**All are biological active!!**

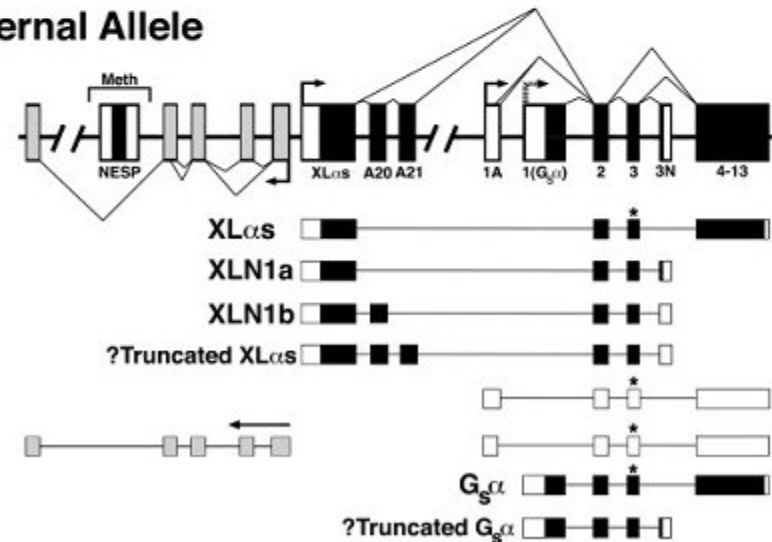
# GNAS locus: role of genetic imprinting

The *Gs* transcripts are **biallelically expressed** in most tissue, but are expressed **primarily from the maternal allele** in some tissue

Maternal Allele



Paternal Allele





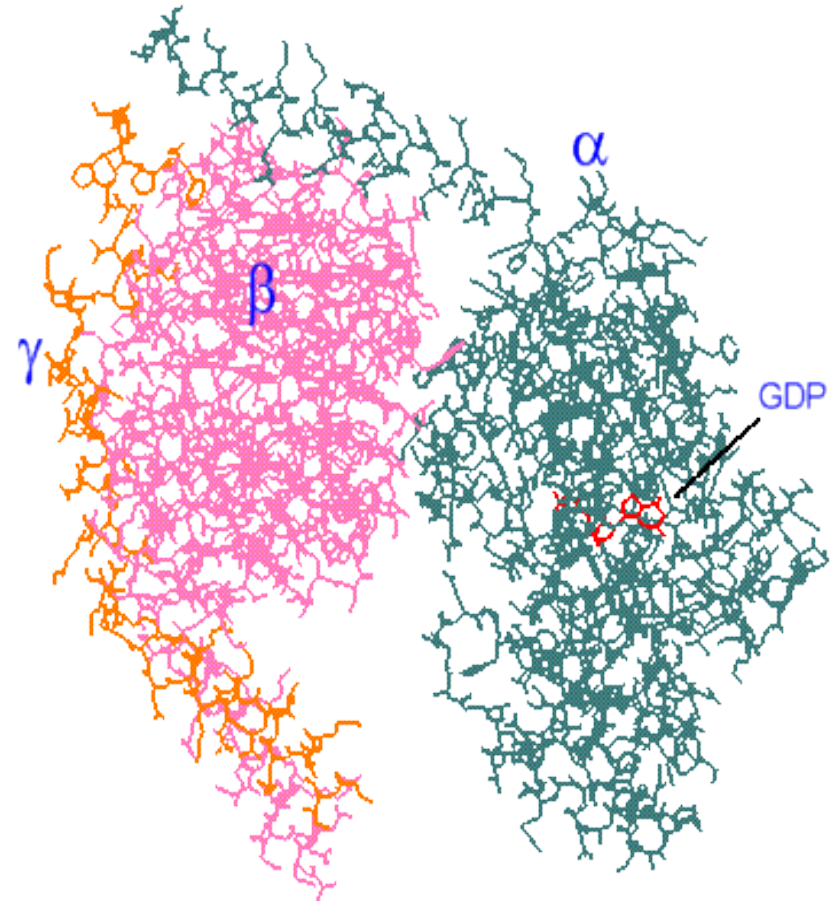
# Gs Proteins

Integral components of diverse signaling pathways:

**ACT AS A MOLECULAR SWITCHES**

Each G protein is defined by:

- specific  $\alpha$ -subunit (39-46 kDa) which binds GDP/GTP
- $\beta$ -subunit (35-39 kDa) and  $\gamma$ -subunit (~ 8 kD) that form a complex

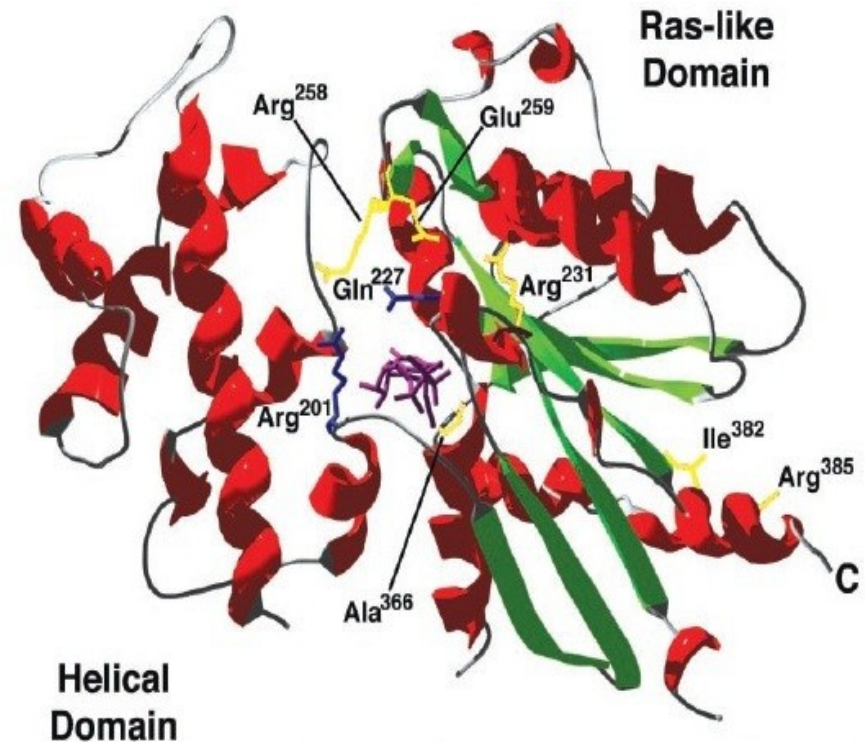


Kobilka et al (1998)

# Gs- $\alpha$ subunit

Binds guanine nucleotide and interacts with specific receptors and effectors

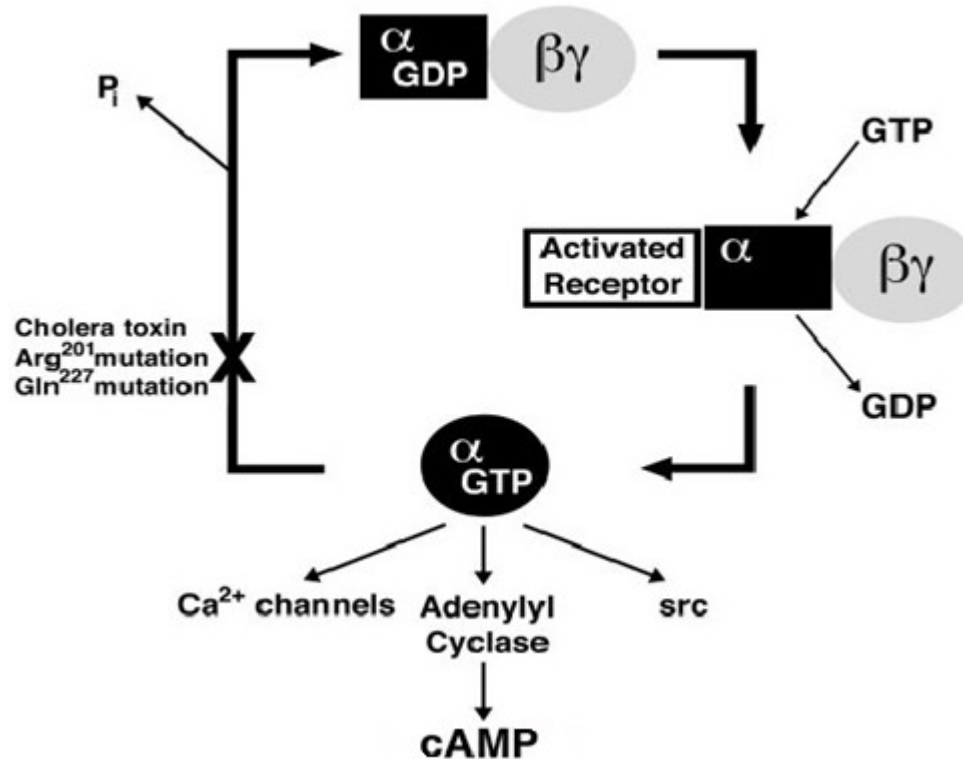
- Mutation of two residues important for the GTPase "turn-off" reaction (Arg201 and Gln227)
- Gsa is a protein found in all cell type except mature spermatozoe
- There are activation and inactivation mutations.



Weinstein et al (2001)

# Gs-a Proteins

Gs is activated and deactivated via the GTPase cycle of its alpha subunit



# Project goal

We are proposing two goals:

1. Create a disease murine models with classic POH
2. Strategy to understand the molecular basis of POH using 3rd generation lentivector

# Project goal

## Brief summary of the procedures:

- creation of two different useful Lentivector to perform our protocols
- check point on the ability of construct
- Use of LV to:
  - generate a mouse model of POH
  - check the efficiency in vitro
- Subsequently, if everything works well, use LV protocol directly on mice

# Gene Therapy strategy: 3rd generation lentivector

## WHY LENTIVECTOR?!

### Advantages:

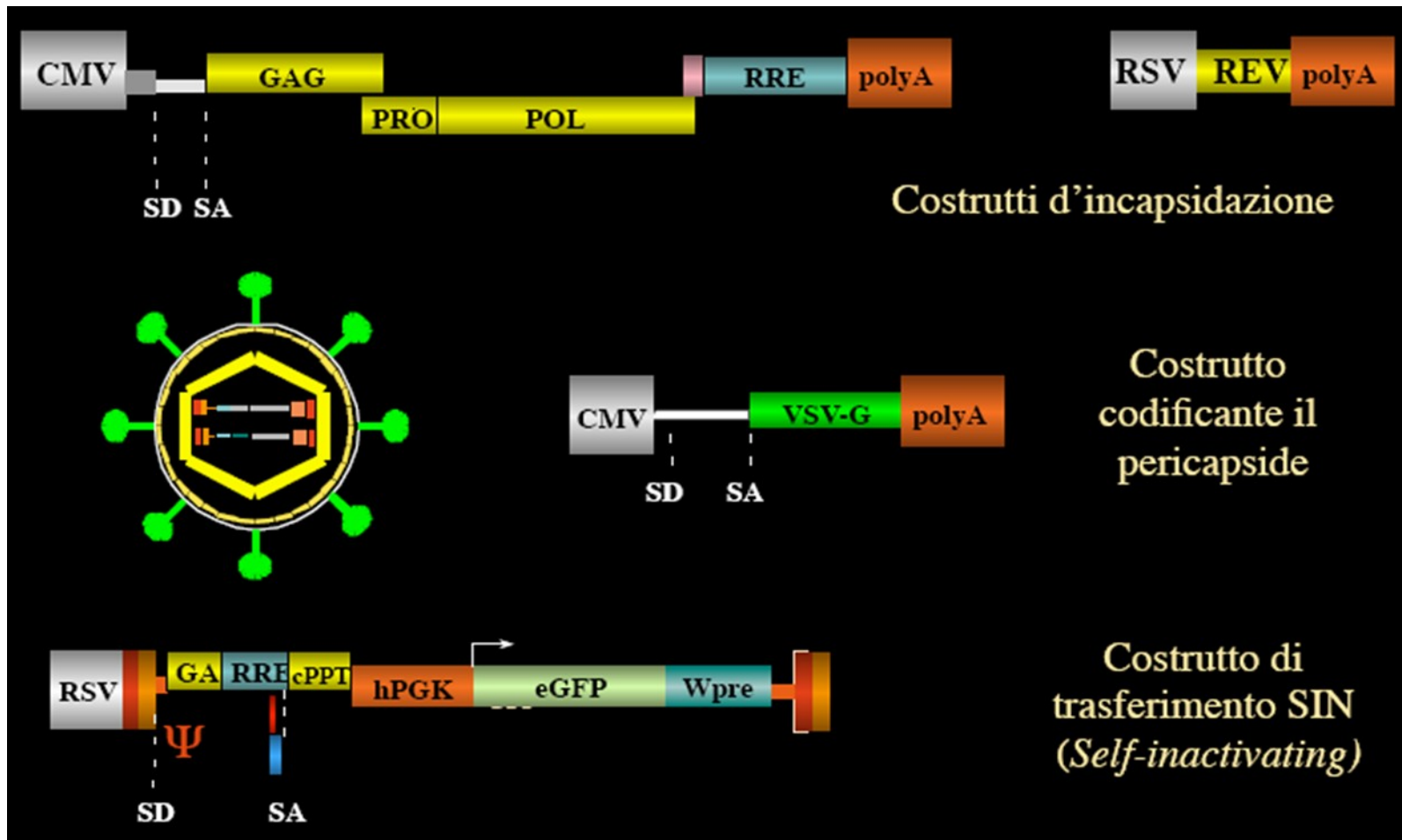
- can accommodate transgene large up to 8 kb
- stable long term transgene expression
- high titers
- easy to prepare

### Drawbacks:

- Derives from the pathogenic HIV-1
- insertional mutagenesis

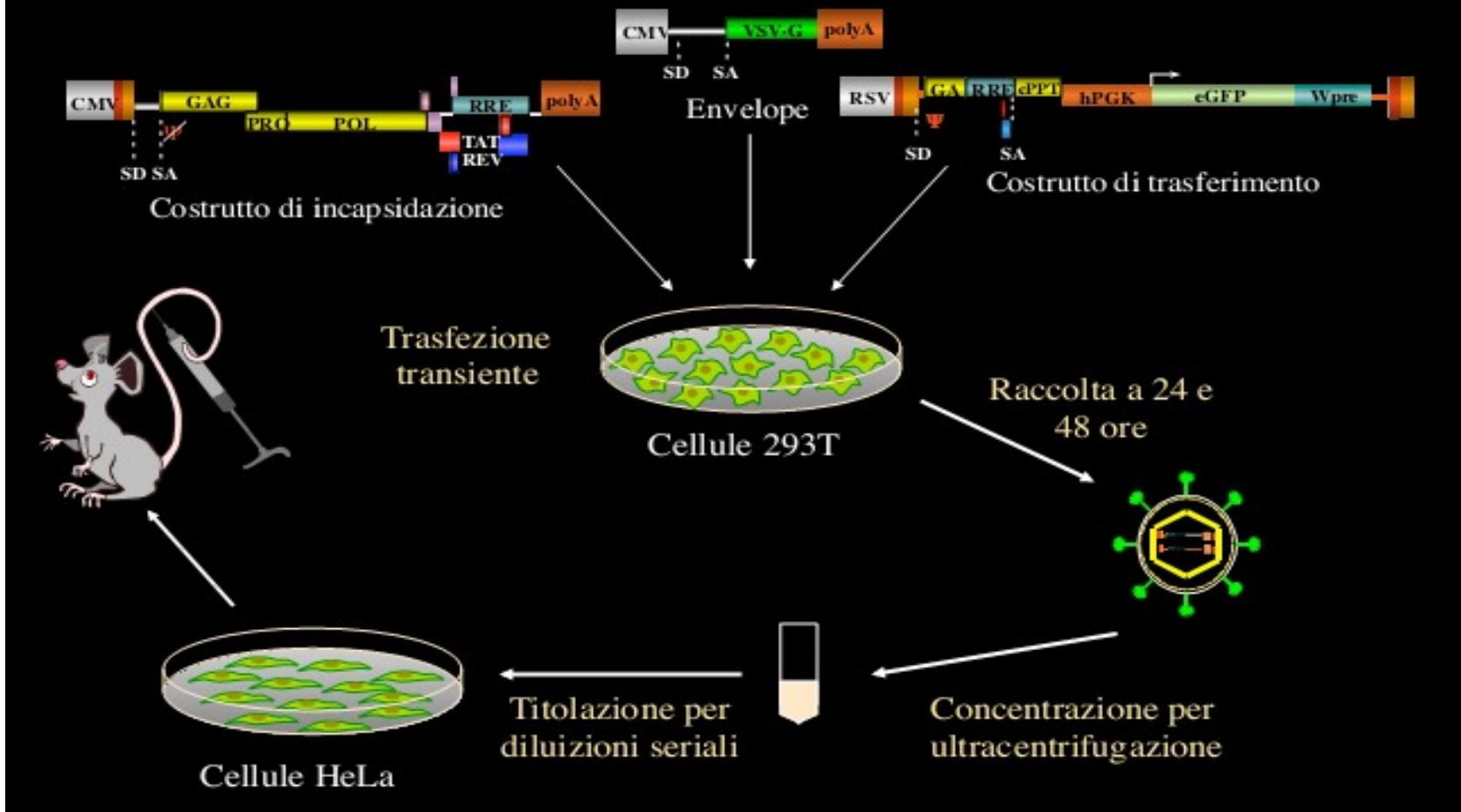
# Gene Therapy strategy: 3rd generation lentivector

## HOW PRODUCE LV IN LAB



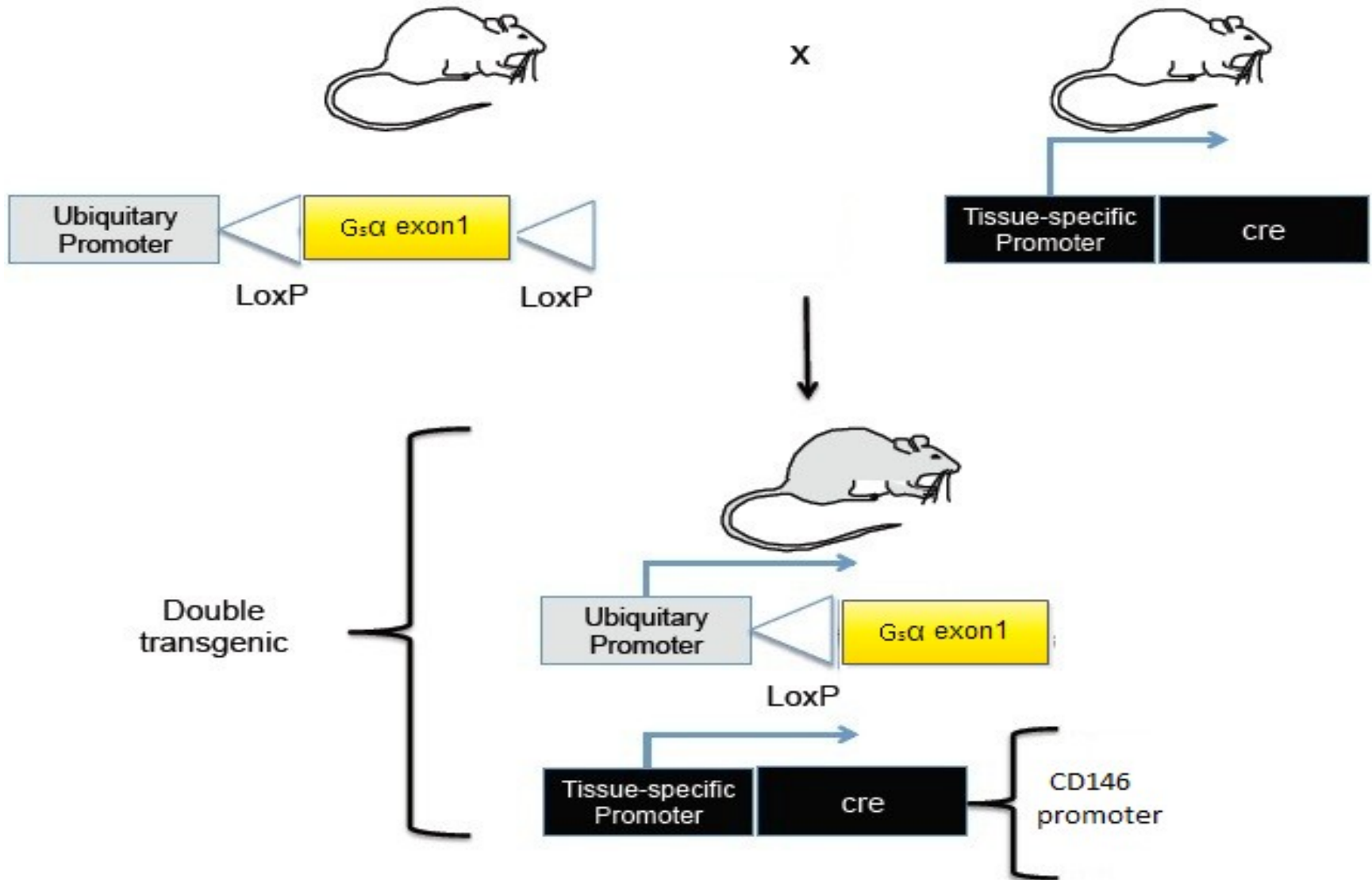
# Gene Therapy strategy: 3rd generation lentivector

## Produzione del vettore



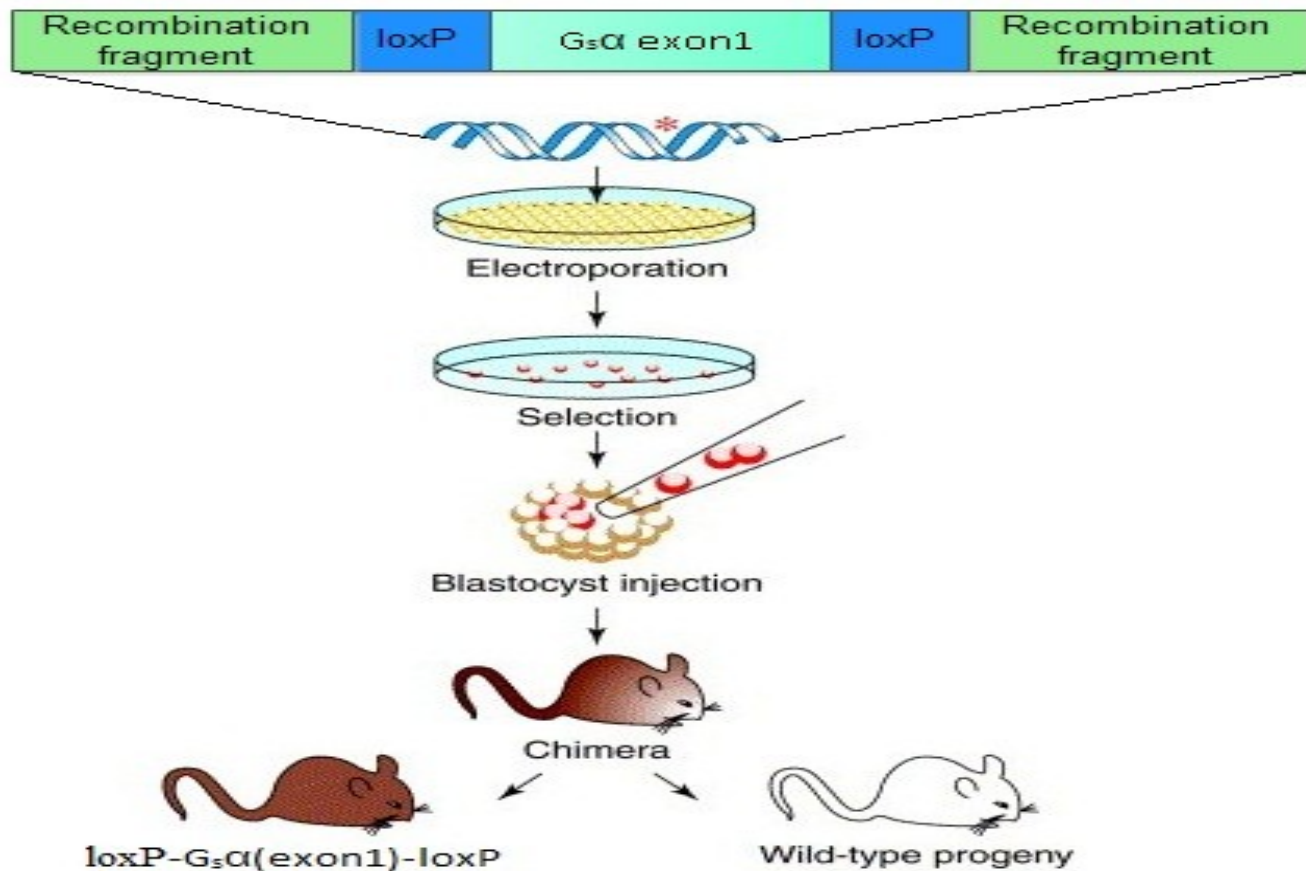


# 1. Murine model for POH



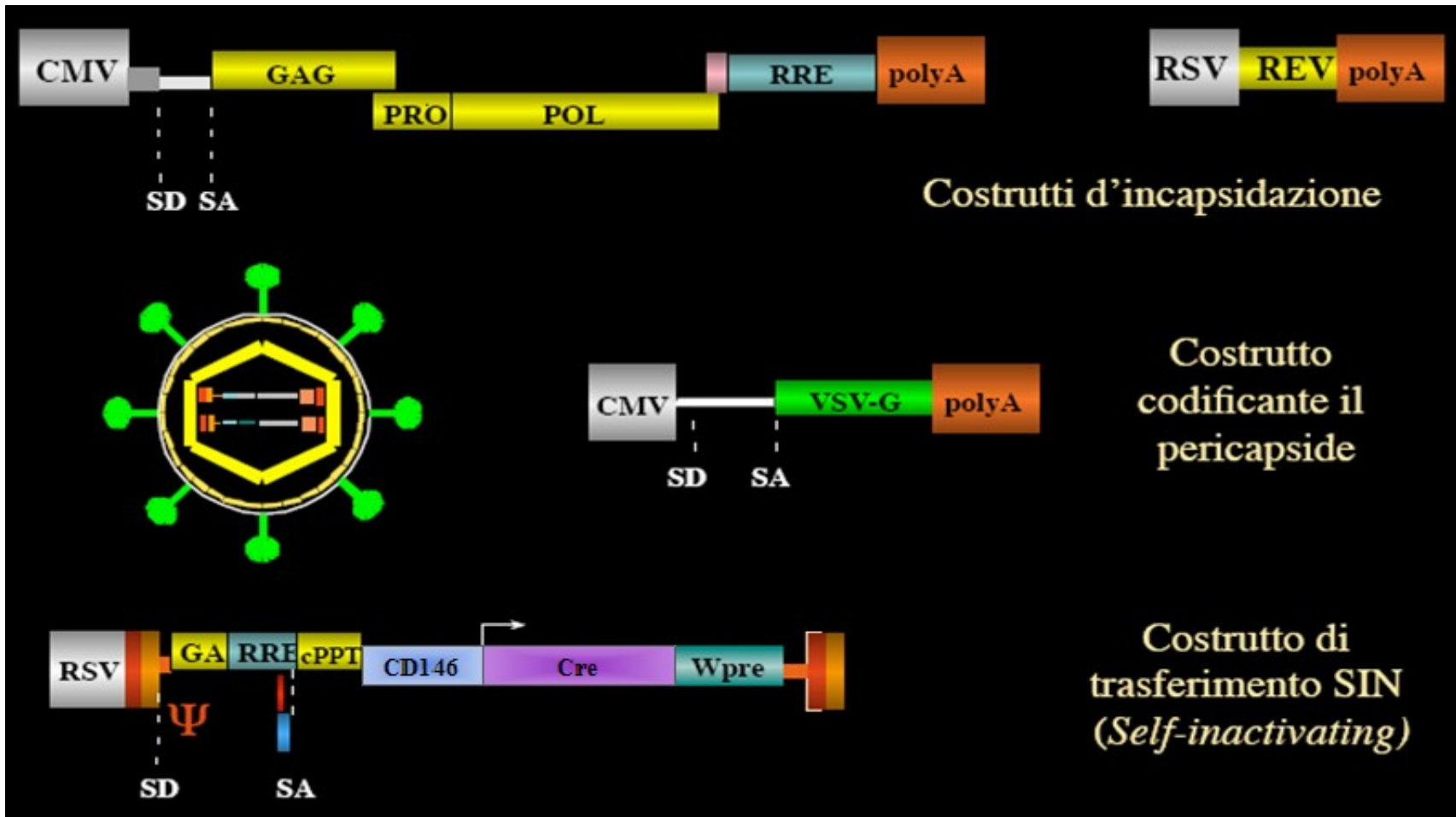
# 1. Murine model for POH

## Generation of loxP-Gs $\alpha$ (exon1)-loxP mouse



# 1. Murine model for POH

## Generation of CD146-Cre mouse



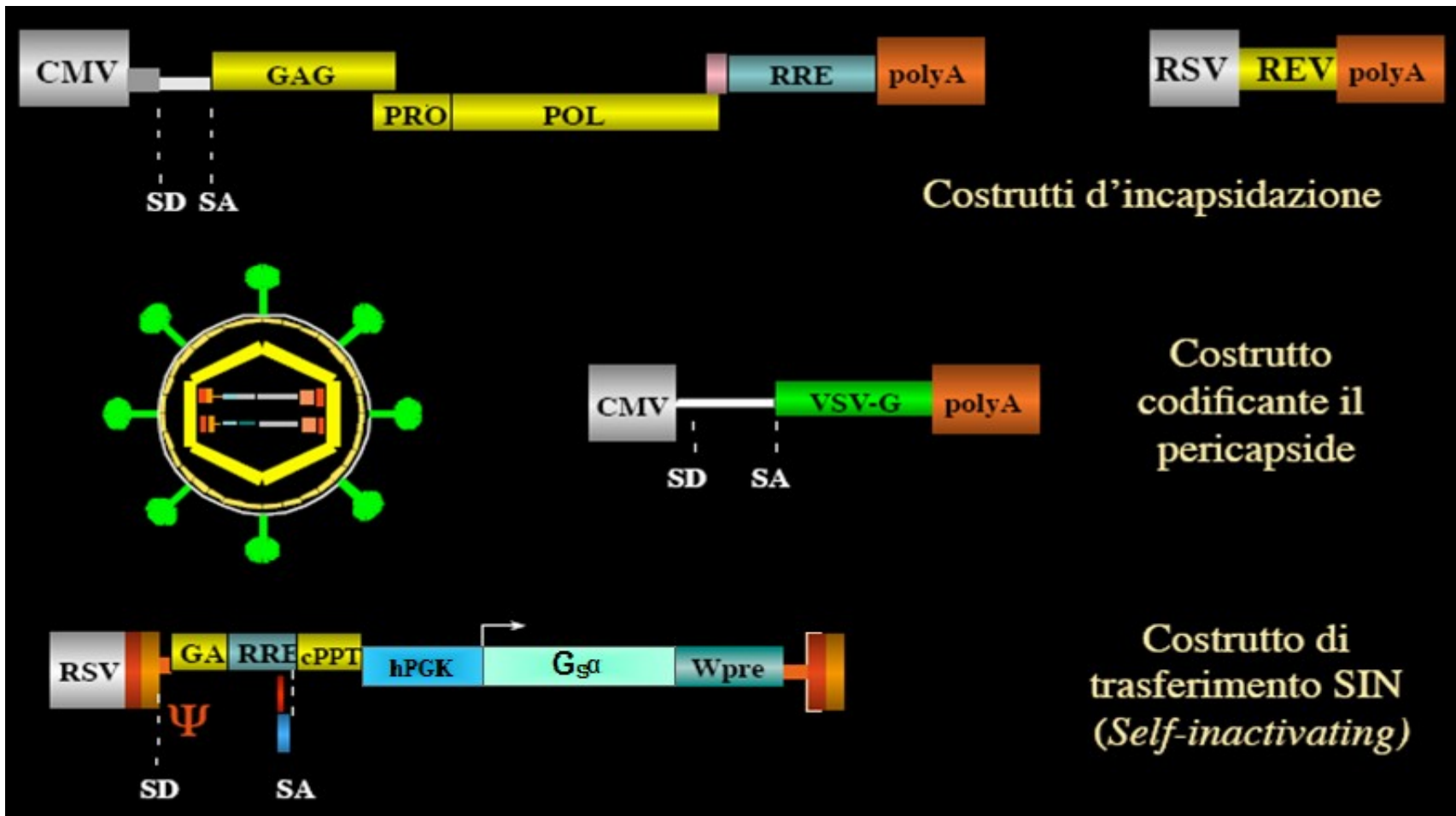
# 1. Murine model for POH

Controls on F1 looking for double transgenic mice:

- Search mutated phenotype
- Quantification of *Gs- $\alpha$*  mRNA and protein (Western blot, RT-PCR ... )

## 2. Gene Therapy strategy: 3rd generation lentivector

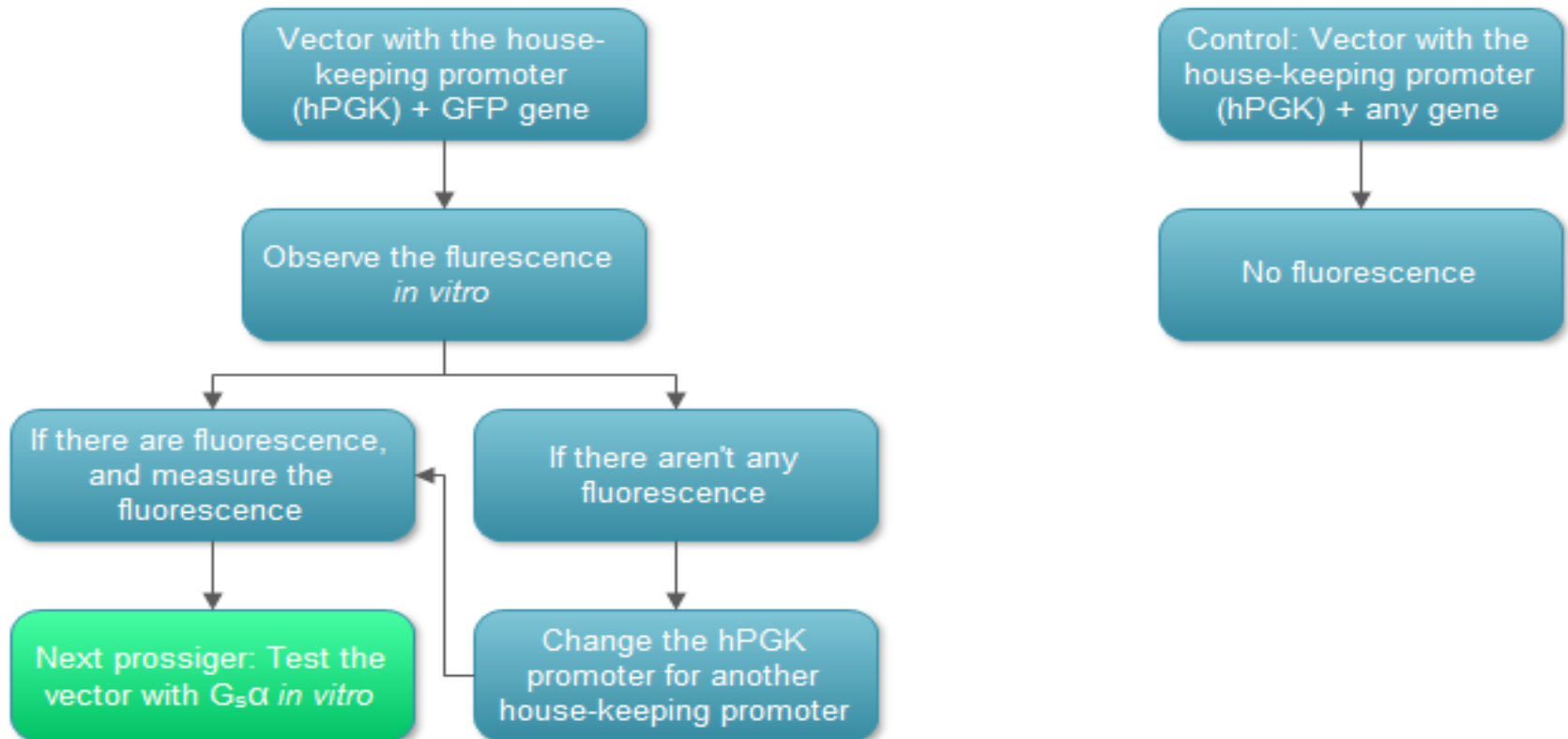
### Production of LV-Gsa<sup>+</sup>



## 2. Gene Therapy strategy: 3rd generation lentivector

Check point on the efficiency of LV-GFP on MSC

Test the efficacy of the vector  
Cell line: wild type mouse's mesenchymal cells



# POH: new model for Gene Therapy using Lentivectors

At this point, we had:

- used a transgenic approach to make animal model (Cre-Lox strategy to have a double transgenic mouse phenotypically and genotypically POH)
- test step by step if the protocols used was right performed
- construct LV- $Gsa^+$
- test if the LV- $Gsa^+$  can be useful to produce a correct form of  $Gsa$  in cells explant from our mouse POH

**Now, where is the connection between  $Gs-\alpha$  mutation and POH?!**

## 2. Gene Therapy strategy

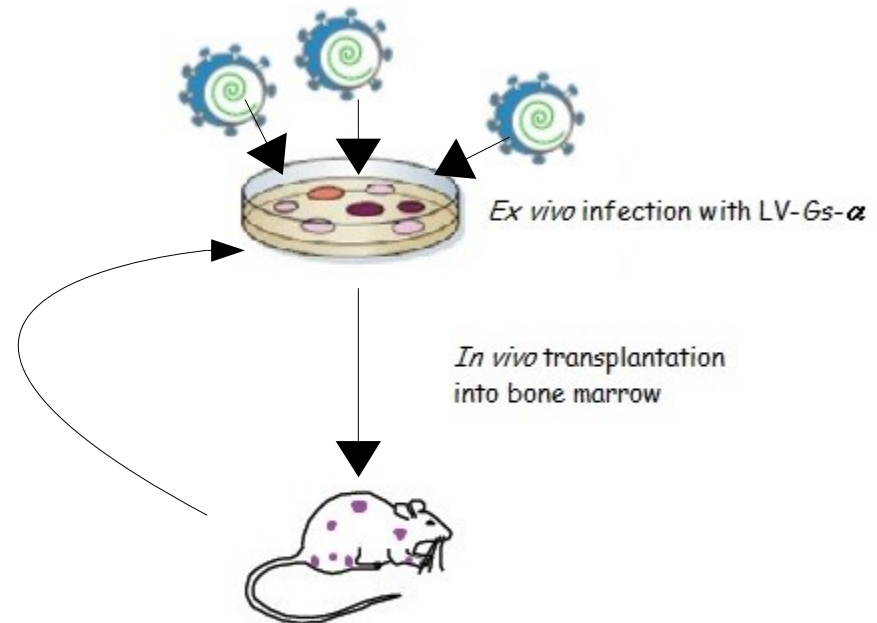
Gs- $\alpha$  loss of function decrease the cAMP and alters the MSC differentiation pathway



## 2. Gene Therapy strategy

### Briefly:

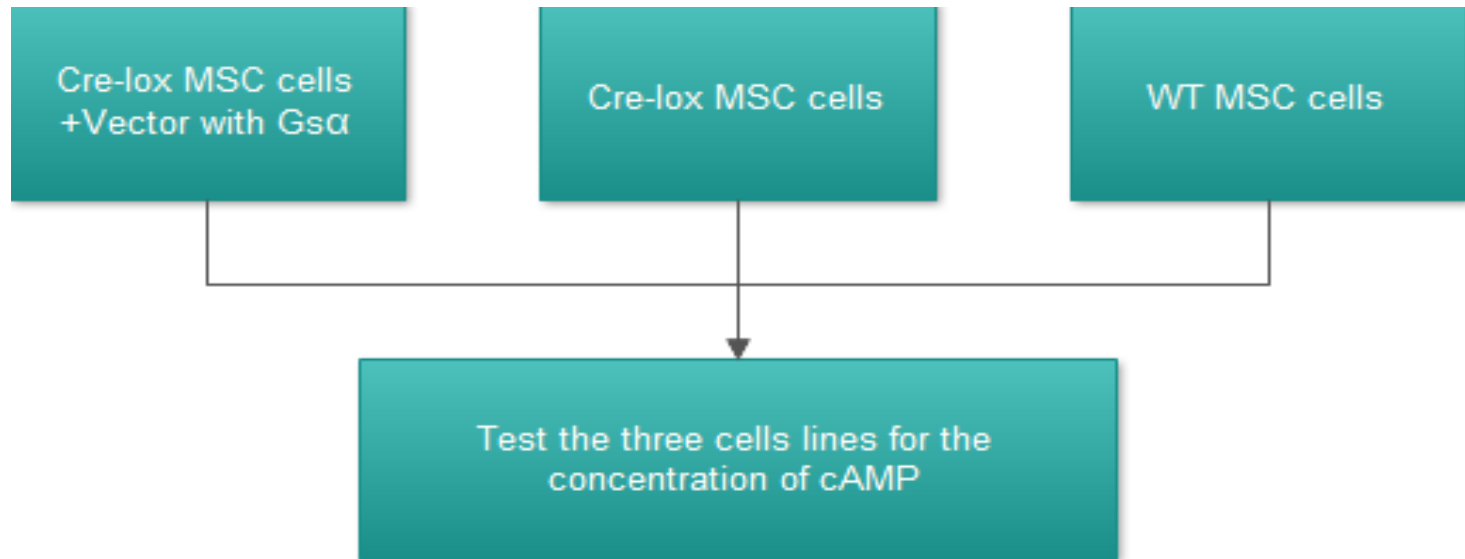
- explantation of MSC
- treatment with LV-Gs- $\alpha$
- transplantation in POH mouse model



## 2. Gene Therapy strategy

But... before go head let's check if it works!

Compared concentration of cAMP in the three cell lines.

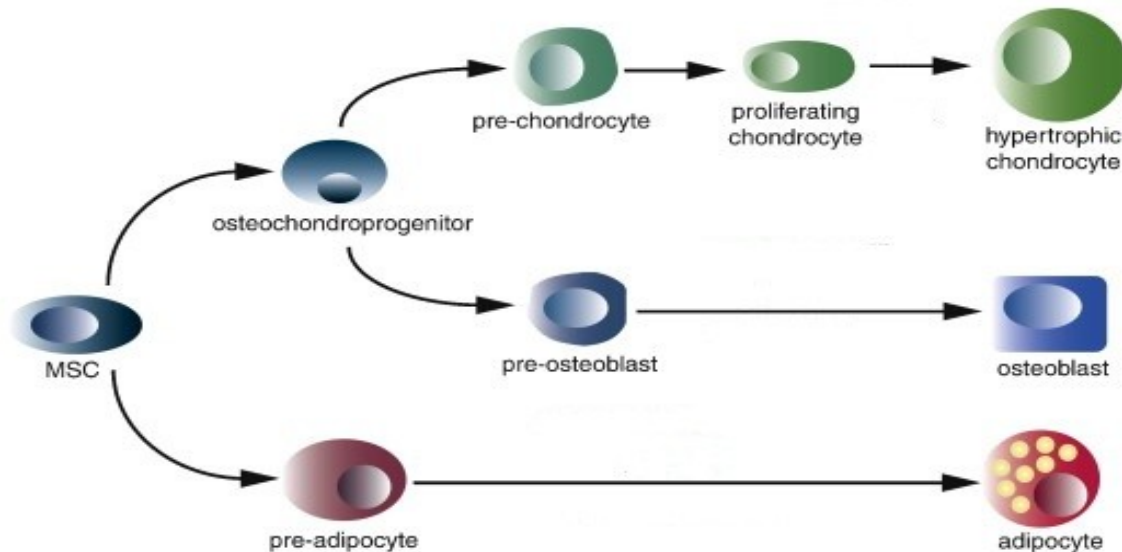


We will measure the concentration of cAMP using a commercial kit assay

## 2. Gene Therapy strategy

### Why target MSC (Mesenchymal Stem Cells)?

- POH patients have disorder of Mesenchymal differentiation
- MSC are the multipotential progenitors of:  
skeletal cells (**osteoblasts**, **chondrocytes**, hematopoietic-supportive stromal cells) and **adipocytes**



Karsenty (2008)

## 2. Gene Therapy strategy

### MSC (Mesenchymal Stem Cells)

#### Properties:

- can be obtained from various tissues (ex: bone marrow, umbilical cord blood (UCB), placental tissue, and adipose tissue)
- isolated on the basis of their adhesive properties ( ex: CD146)
- *in vitro*:
  - > differentiation is initiated by addition of growth factors and low molecular weight components
  - > characterized by their ability to adhere to plastic in culture and differentiate into various mesodermal cell lineages

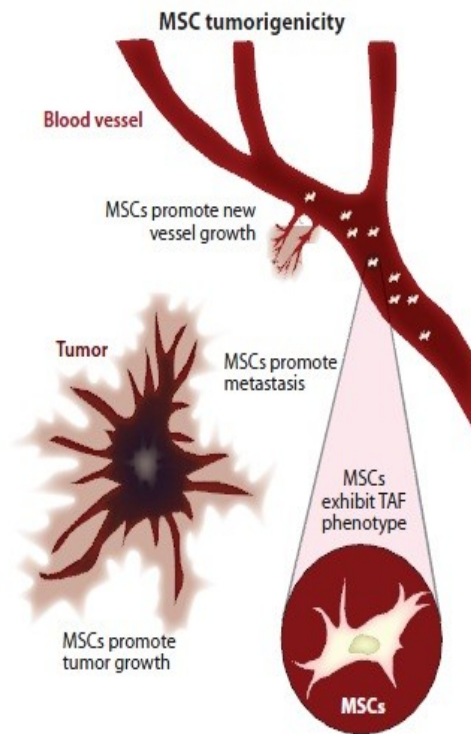
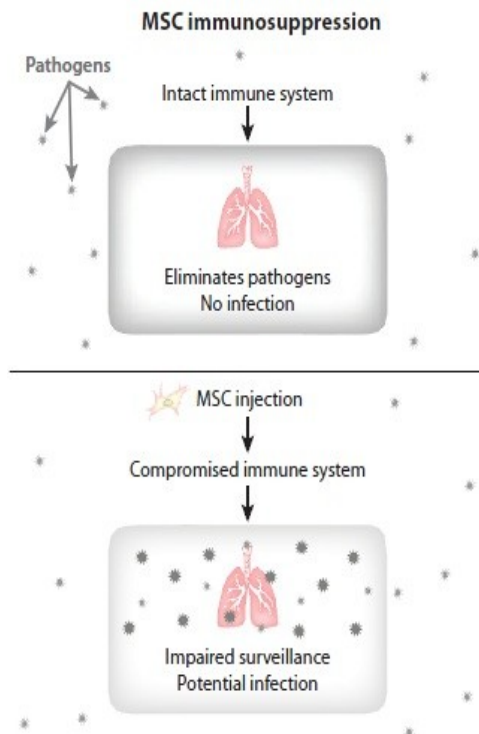
**MSC transplantation represents an exciting approach that could potentially treat complex diseases by providing combinatorial therapy!**

# 2. Gene Therapy strategy

## MSC (Mesenchymal Stem Cells)

### Drawbacks:

- recent preclinical studies have highlighted potential long-term risks include potential maldifferentiation, immunosuppression, and instigation of malignant tumor growth



- lack of knowledge to conclusively:

- lineage relationships
- differentiation properties
- no culture conditions have been described which can maintain multipotency over time

**Nature of these cells is poorly understood!**

# Expected Results

Increase the knowledge on the  
molecular  
mechanisms by which POH is caused,  
allow to schedule a targeted therapy.

# Future Prospectives

Based on new data, be able to allow in the future to implement our strategy directly on

POH patients

# Technics

- Tissue-specific expression (transgene expression, cAMP test, downstream effectors)
- RX analysis
- Histological analysis
- Histomorphometry
- Ex vivo differentiation assay (osteo/adipo)
- Osteoclastogenesis assay
- Molecular analysis
- .....



# Material and Costs

- PCR: 1000 €
- Cloning kit: 700 €
- Lentiviral vector : 650 €
- Transgenic mouse 200 €/ month
- Mice floxed: 1000 €
- RT-qPCR: 250 €
- Western blot: 200 €
- Molecular analyse (antibodies, reagent etc ) 2000 €
- Discartable material 1000 €
- cAMP kit commercial kit assay 325 €

 invitrogen™

 celectis  
bioresearch



We are supposing to spend around 16500€ to perform our project in 5 years

# References

- Bianco, P., et al (2008). Skeletal progenitors and the *GNAS* gene: fibrous dysplasia of bone read through stem cells. **Endocrine Reviews** 22(5):675-705.
- Kaplan, M., et al. (2002). Paternally inherited inactivating mutations of the *GNAS1* gene in Progressive Osseous Heteroplasia. **N Engl J Med** Vol. 346, No. 2.
- Kaplan, M., et al (2000). Progressive Osseous Heteroplasia. **Journal of bone and mineral research** 15:2084-2094.
- Lowe, W.L., et al (2010). G Protein Coupled Receptors in Embryonic Stem Cells: A Role for Gs-Alpha Signaling. **PLoS ONE** 5: e9105.
- Bonthron, D.T., et al (1998) The human *GNAS1* gene is imprinted and encodes distinct paternally and biallelically expressed G proteins. **PNAS** 95:10038-10043.
- Nissenson, R.A. (2012). Gs-a in osteoblasts regulates bone formation and osteoblast differentiation. **IBMS BoneKEy** number 91.
- Otte, A., et al (2012). Mesenchymal stem cells as all-round supporters in a normal and neoplastic microenvironment. **Cell Communication and Signaling** 10:26.
- Ghannam, S., et al (2010). Immunosuppression by mesenchymal stem cells: mechanisms and clinical applications. **Stem Cell Research & Therapy** 1:2.
- Kottler, M.L., et al (2010). Progressive Osseous Heteroplasia: A Model for the Imprinting Effects of *GNAS* Inactivating Mutations in Humans. **J Clin Endocrinol Metab** 95(6):3028-3038.
- Tsumaki, N., et al (2006). Bone Morphogenetic Proteins in Bone Stimulate Osteoclasts and Osteoblasts During Bone Development. **Journal of bone and mineral research** 21:7.

# References

- Nissenson, R., et al (2012). Cyclic AMP signaling in bone marrow stromal cells has reciprocal effects on the ability of mesenchymal stem cells to differentiate into mature osteoblasts versus mature adipocytes. **Endocrine** 42:622-636.
- Klein, G., et al (2011). Regeneration of cartilage and bone by defined subsets of mesenchymal stromal cells—Potential and pitfalls. **Advanced Drug Delivery Reviews** 63:342–351.
- McTaggart, S.J., et al (2008). Immunosuppression by mesenchymal stromal cells: From culture to clinic. **Experimental Hematology** 36:733-741.
- Karsenty, G., et al (2009). A paradigm of integrative physiology, the crosstalk between bone and energy metabolisms. **Molecular and Cellular Endocrinology** 310:21-29.
- Bonnet, D., et al (2011). Prospective identification and isolation of murine bone marrow derived multipotent mesenchymal progenitor cells. **Best Practice & Research Clinical Haematology** 24:13–24.
- Bellocq, J.P., et al (2000). Progressive osseous heteroplasia: an uncommon cause of ossification of soft tissues. **Annales de Génétique** 43:75-80.
- Chan, I., et al (2004). Progressive osseous heteroplasia resulting from a new mutation in the *GNAS1* gene. **Clinical and Experimental Dermatology** 29:77-80.
- Weinstein, L.S., et al (2004). Minireview: *GNAS*: Normal and Abnormal Functions. **Endocrinology** 145(12):5459-5464.
- Weinstein, L.S., et al (2001). Endocrine Manifestations of Stimulatory G Protein  $\alpha$ -Subunit Mutations and the Role of Genomic Imprinting. **Endocrine Reviews** 22(5):675-705.
- Nardi, N.B., Et al (2003). Murine marrow-derived mesenchymal stem cell: isolation, in vitro expansion, and characterization. **British Journal of Haematology** 123:702-711.

# References

- Han, J., et al (2012). Therapeutic application of mesenchymal stem cells in bone and joint diseases. **Clin Exp Med**.
- Milwid, J.M., et al (2010). Mesenchymal Stem Cells as Therapeutics. **Annu. Rev. Biomed. Eng.** 12:87-117.
- Kaziro, Y., et al (1988). Isolation and characterization of the human *Gsa* gene. **Proc. Natl. Acad. Sci.** 85:2081-2085.
- Pignolo, R.J., et al (2008). Diagnostic and Mutational Spectrum of Progressive Osseous Heteroplasia (POH) and Other Forms of *GNAS*-based Heterotopic Ossification. **Am J Med Genet A.** 146A(14):1788-1796.
- Murat, B. (2007). The *GNAS* Locus: Quintessential Complex Gene Encoding *Gsa*, *XLas*, and other Imprinted Transcripts. **Current Genomics** 8:398-414.
- Kessler, J.A., et al (2010). Animal Models of typical heterotopic ossification. **Journal of Biomedicine and Biotechnology** ID 309287
- Kronenberg, H.M., et al (2011). *Gsa* enhances commitment of mesenchymal progenitors to the osteoblast lineage but restrains osteoblast differentiation in mice. **The Journal of Clinical Investigation** 21.
- Weinstein, L.S., et al (2009). *GNAS* haploinsufficiency leads to subcutaneous tumor formation with collagen and elastin deposition and calcification. **Endocr Res.** 34(1): 1-9.
- Ferrari, P., et al (2008). Eteroplasia Ossificante Progressiva: una nuova mutazione del gene *GNAS1* e reviews della letteratura. **Medico e Bambino** 11(3).



Waiting for...next season!!!